

Setting Your Own Stain Panels on the FACScalibur:

(You will have to do this for each individual cytometer.)

- Under ACQUISITION choose EDIT PANELS
- Click on NEW...
- Name new panel - example "bea" This will contain ALL my stain combinations.
- HIGHLIGHT the name of the panel you want to edit.
- Click on ADD - a tube will be entered in the middle column as "tube #X". change name if desired - example "unstained"
- HIGHLIGHT the tube you want to enter stain combinations for. Now the right-most column will be activated.
- Fill in stains:
 - p1: FSC- height
 - p2: SSC - height
 - p3: FL1-height
 - p4: FL2-height
 - p5: FL3-height
 - p6: SKIP
 - p7: FL4-height

Remember - P1 & P2 are almost always FSC & SSC and Cy5/APC is P7 - NOT P6!!!

- Click on "ADD" This will add another tube in the middle column.
- Click on "tube #2" - change tube name to "anti-tubulin", then highlight and enter stains in the right column.
 - p1: FSC- height
 - p2: SSC - height
 - p3: anti-tubulin-FITC
 - p4: FL2-height
- Click on ADD - change name - "isotype controls"
 - p1: FSC- height
 - p2: SSC - height
 - p3: IgG2b-FITC
 - p4: IgG1-PE
- Click on ADD - change tube name to "T1"
 - p1: FSC- height
 - p2: SSC - height
 - p3: anti-IL2-FITC

- p4: anti-IFN γ -PE

ADD Infinitum!

When you are setting up your acquisition screen chose your panel in the Parameter Description Window (under Panels - not Default panel) and thereafter select the stain combination for each sample using the pull-down menu . NOTE - the cytometer will automatically advance to the next stain combination after each saved file. Keep this in mind as you enter tubes (I don't know of a way to edit the order of tubes once entered into the panel) and when you are collecting your data.